

# The isolation and identification of useful bacteria that decrease nitrous oxide emission from agricultural field

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## Abstract

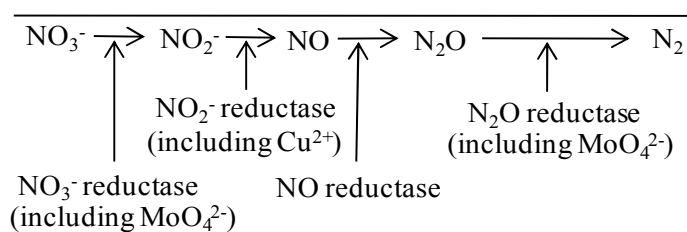
Increases in atmospheric concentration of nitrous oxide (N<sub>2</sub>O) are partly attributed to soil denitrification. To overcome the problem, we propose utilising useful bacteria *in situ* to suppress N<sub>2</sub>O emission from agricultural soil by inhibiting the activities of microorganisms that produce high levels of N<sub>2</sub>O. The useful bacteria are the bacteria that convert nitrate ion (NO<sub>3</sub><sup>-</sup>) to another nitrogen compound under aerobic conditions, or aerobic denitrifying bacteria that reduce NO<sub>3</sub><sup>-</sup> to nitrogen under aerobic condition. Therefore, we attempted screening to find bacteria that have either of those properties, and then, four strains (K, N-I, N-II, and N-III) that could remove NO<sub>3</sub><sup>-</sup> and NH<sub>4</sub><sup>+</sup> under aerobic conditions were isolated and identified. Of the total nitrogen originally provided, 55% to 75% was taken up into the cells and the residual nitrogen went into the culture supernatant. The strains K, N-I, and N-II were identified as *Enterobacter cloacae*. The strain N-III showed 98.2% gene sequences identical to *Enterobacter cloacae* and 98.1% to *Klebsiella pneumonia*, indicating that the strain N-III has the potential to be a new species of bacterium.

## Key Words

Aerobic denitrifying bacteria, identification, nitrous oxide.

## Introduction

Nitrous oxide (N<sub>2</sub>O) has a 200- to 300-fold-stronger greenhouse effect than carbon dioxide (CO<sub>2</sub>). It has been reported that N<sub>2</sub>O has the potential to destroy the ozone layer (Takaya *et al.* 2003). Recently, the concentration of N<sub>2</sub>O in atmosphere is increasing. One of the proposed sources of N<sub>2</sub>O is soil denitrification of nitrogenous compounds (Figure 1), resulting from excess agricultural fertilizer. The nitrate ion by fertilization is reduced by denitrifying bacteria to gaseous nitrogen (N<sub>2</sub>) under anaerobic condition. However, the agricultural field is aerobic, so the reduction of N<sub>2</sub>O to N<sub>2</sub>, that is, the final step of denitrification, is suppressed by oxygen. Therefore, considerable amounts of N<sub>2</sub>O are released in agricultural fields. To decrease N<sub>2</sub>O emission, we propose dispersing useful bacteria over the agricultural field where the aerobic denitrifiers grow and suppress activities of microorganisms that produce high levels of N<sub>2</sub>O. The useful bacteria we propose convert NO<sub>3</sub><sup>-</sup> to another nitrogen compound or denitrify releasing low levels of N<sub>2</sub>O under aerobic condition. Therefore, we attempted the isolation and identification of bacteria that have either of those properties.



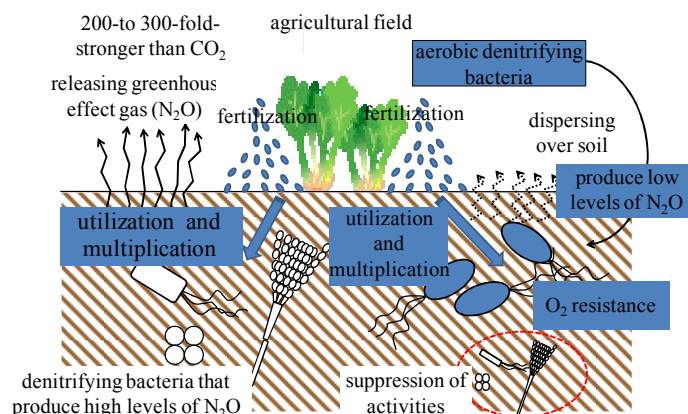
**Figure 1.** The given course of denitrification steps.

## Materials and Methods

### Isolation of aerobic denitrifiers

At first, we collected 77 samples of rhizosphere soil from agricultural fields in Kurokawa, Machida, and Nishinomiya (Japan), and then transferred to them saline (0.8%) to adjust the suspensions. Some drops of the suspensions were transferred to 5 mL of screening media in flasks with cotton plugs, respectively and cultured by shaking at 130 rpm at 30 °C. The screening medium contained the following compounds: 0.1% NH<sub>4</sub>NO<sub>3</sub>, 2% Glucose (Glc), and trace metallic salts. Two of the trace metallic salts, molybdate ion (MoO<sub>4</sub><sup>2-</sup>) and copper ion (Cu<sup>2+</sup>), are needed for enzymes to catalyze some steps of denitrification (reduction of NO<sub>3</sub><sup>-</sup> to NO<sub>2</sub><sup>-</sup>, of NO<sub>2</sub><sup>-</sup> to NO, and of N<sub>2</sub>O to N<sub>2</sub>) (Figure 2). Bacterial growth, consumption of NO<sub>3</sub><sup>-</sup>, NH<sub>4</sub><sup>+</sup> (N-

compounds), and Glc and production of  $\text{NO}_2^-$  were monitored by quantitative analysis. After some independent experiments as described previously, positive strains were isolated and cultured further in 3 mL of the same media at 130 rpm at 30 °C, to study the properties of isolated strains.



**Figure 2. Suppression of releasing  $\text{N}_2\text{O}$  by aerobic denitrifying bacteria.**

#### *Analysis of total nitrogen concentration in cultural supernatant and cells*

Total nitrogen in the culture supernatant and the cells was determined by Kjeldahl apparatus.

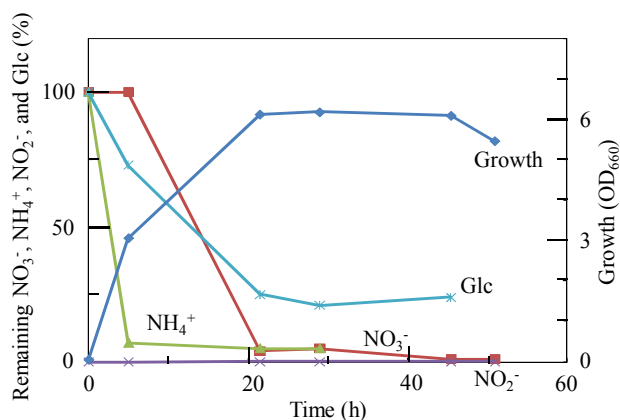
#### *Identification of the strains*

The 16S rRNA genes (1500 bp) from the total DNA extracted from the four strains were amplified by PCR. The PCR products were ligated into a pGEM-T easy vector and transformed into *Escherichia coli* cells and then, clones were sequenced. Sequence results were compared with reference sequences using DNA Data Bank of Japan (DDBJ) service.

## Results

#### *Isolation of aerobic denitrifiers*

It was found that four strains of 77 samples have the potential to be aerobic denitrifiers and we defined the four strains as K, N- I , N- II , and N-III. N- II was able to remove more  $\text{NO}_3^-$  and  $\text{NH}_4^+$  than other three strains, so the result of N- II was shown in Figure 3. The remaining  $\text{NO}_3^-$ ,  $\text{NH}_4^+$ , and Glc in the medium decrease sharply for the first 20 hours. The production of  $\text{NO}_2^-$  was too little to detect. Although  $\text{NO}_3^-$  and  $\text{NH}_4^+$  were almost removed, the value of growth was under 6 (optical density at 660 nm [ $\text{OD}_{660}$ ]). This result may indicate that  $\text{NO}_3^-$  is not being assimilated because this would expect a growth value much higher than 6. To further study the nitrogen prevalence, the four strains were cultured in 50 mL of same media under same conditions. After  $\text{NO}_3^-$ ,  $\text{NH}_4^+$  were removed, to analyze total nitrogen, the cultural supernatant and cells were collected.



**Figure 3. Growth profile of strain N- II and of N-compounds in the medium with  $\text{MoO}_4^{2-}$  and  $\text{Cu}^{2+}$  consumption**

#### *Analysis of total nitrogen in cultural supernatant and cells*

Table 1. shows utilization of 0.1%  $\text{NH}_4\text{NO}_3$  (initially 1.24 mmol) and total nitrogen. *Paracoccus denitrificans* and *Pseudomonas stutzeri* were used as controls in that they are known to denitrify releasing

low level of  $\text{N}_2\text{O}$  under aerobic conditions. The controls removed almost all  $\text{NH}_4^+$  but did not remove  $\text{NO}_3^-$ . With the four isolated strains, 58% to 82% of the total nitrogen originally provided was observed in the cells and 18% to 42% of total residual nitrogen was in the cultural supernatants. Considering remaining  $\text{NH}_4^+$  and  $\text{NO}_3^-$ , it was found that the supernatants contained another nitrogen compound.

**Table 1. Total nitrogen in cultural supernatant and cells**

| strain                          | Initial (mmol ) |       | Finish (mmol )           |                           | remaining rate (%) |
|---------------------------------|-----------------|-------|--------------------------|---------------------------|--------------------|
|                                 | Sup.            | Cells | Sup. ( $\text{NO}_3^-$ ) | ( $\text{NH}_4^+$ ) Cells |                    |
| K                               | 1.24            | 0     | 0.60 (0.21)              | (0.04) 0.79               | 112                |
| N-I                             | 1.24            | 0     | 0.57 (0.15)              | (0.04) 0.75               | 106                |
| N-II                            | 1.24            | 0     | 0.33 (0.00)              | (0.03) 1.02               | 109                |
| N-III                           | 1.24            | 0     | 0.59 (0.14)              | (0.03) 0.73               | 106                |
| <i>Paracoccus denitrificans</i> | 1.24            | 0     | 1.00 (0.58)              | (0.04) 0.48               | 119                |
| <i>Pseudomonas stutzeri</i>     | 1.24            | 0     | 1.08 (0.67)              | (0.05) 0.41               | 121                |

#### Identification of the strains

All of the identification results of four strains are shown in Table 2. The strain K showed 99.0% identity to *Enterobacter cloacae* isolate 766 and 98.4% identity to *Enterobacter cloacae* B5. The strain N- I showed 99.1% identity to *Enterobacter cloacae* strain FR and 98.6% identity to *Enterobacter cloacae* isolate 766. The strain N- II showed 98.6% identity to *Enterobacter cloacae* isolate 766 and 98.0% identity to *Enterobacter cloacae* strain B5. The three strains had significantly-high identity to *Enterobacter cloacae*. The strain N-III showed 98.2% identity to *Enterobacter cloacae* isolate 766 and 98.1% identity to *Klebsiella pneumoniae* strain TCCC1, indicating that N-III does not have significantly-high identity.

**Table 2. The identification of strains K, N- I , N- II , and N-III**

| Isolate | Homologous strains                        | Identity (%) |
|---------|---|--------------|
| K       | <i>Enterobacter cloacae</i> isolate 766   | 99.0         |
|         | <i>Enterobacter cloacae</i> strain B5     | 98.4         |
| N-I     | <i>Enterobacter cloacae</i> strain FR     | 99.1         |
|         | <i>Enterobacter cloacae</i> isolate 766   | 98.6         |
| N-II    | <i>Enterobacter cloacae</i> isolate 766   | 98.6         |
|         | <i>Enterobacter cloacae</i> strain B5     | 98.0         |
| N-III   | <i>Enterobacter cloacae</i> isolate 766   | 98.2         |
|         | <i>Klebsiella pneumoniae</i> strain TCCC1 | 98.1         |

## Conclusions

#### Isolation of aerobic denitrifiers

Four strains removed  $\text{NO}_3^-$  and  $\text{NH}_4^+$  under aerobic conditions but by analysis of total nitrogen in the supernatant and cells, denitrification did not occur. Considering the remaining  $\text{NH}_4^+$  and  $\text{NO}_3^-$ , it was postulated that the supernatants contained another nitrogen compound.

#### Identification of the strains

The strains K, N- I , and N- II were identified as *Enterobacter cloacae*. The nucleotide sequence of 16S rRNA of N-III could not be identified with known sequences, so we conclude N-III has the potential to be a new species of bacterium.

## Acknowledgements

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## References

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